In vivo deep brain imaging of rats using oral-cavity illuminated photoacoustic computed tomography

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Abstract. Using internal illumination with an optical fiber in the oral cavity, we demonstrate, for the first time, photoacoustic computed tomography (PACT) of the deep brain of rats in vivo. The experiment was performed on a full-ring-array PACT system, with the capability of providing high-speed cross-sectional imaging of the brain. Compared with external illumination through the cranial skull, internal illumination delivers more light to the base of the brain. Consequently, in vivo photoacoustic images clearly reveal deep brain structures such as the hypothalamus, brain stem, and cerebral medulla.

Keywords: oral-cavity illumination; internal illumination; deep brain imaging; photoacoustic computed tomography; optical fiber; anatomic imaging.

Paper 140725R received Nov. 3, 2014; accepted for publication Jan. 6, 2015; published online Jan. 22, 2015.
Intensities at 45 deg and 90 deg were, respectively, \(\sim 56\%\) and \(\sim 15\%\) of the intensity at 0 deg [Fig. 1(b)], where the pinhole faces the reflecting surface of the prism directly and the light intensity is the highest. The maximum light intensity at the palate surface was approximately \(20 \text{ mJ/cm}^2\), which was below the American National Standards Institute (ANSI) limit (29 mJ/cm²) at the chosen wavelength. The 90-deg bent portion of the fiber was wrapped in aluminum foil to prevent light leakage. The total energy at the output end of the fiber was \(\sim 10 \text{ mJ}\). To facilitate fiber insertion through the mouth, the fiber tip was lubricated with water-based lubricant (K-Y® ultra-gel). The insertion was smooth and should not cause any pain that might alter the neural activity.

The system also has a flippable mirror in front of the laser to alternatingly guide the light to the top of the rat’s head for external illumination as a control [Fig. 1(a)]. The redirected light first reached an optical diffuser (EDC-10, RPC Photonics) for homogenization, and then illuminated the rat scalp from above. The light intensity was approximately \(10 \text{ mJ/cm}^2\) at the scalp surface, also below the ANSI limit. The total energy incident on the animal was \(\sim 28 \text{ mJ}\). The flippable mirror design allows easy switching between internal and external illuminations.

Before the experiment, the hair on the head of the rat was removed with depilatory lotion. As shown in Fig. 1(a), the rat holder consisted of a plastic base and a plastic bar as a chin holder. The rat was mounted in an upright position and was secured to the holder with paper tapes (not shown in the sketch). The bent fiber was then inserted into the rat’s oral cavity and also fixed on the holder with paper tapes. We then mounted the holder on a translation stage for elevational scans. The rat was placed underneath the water tank with its scalp coupled to the flexible membrane through ultrasonic gel (Aquasonic, Parker Laboratories, Inc.) to be imaged through the membrane.

The PACT system utilized a 5-MHz (80% bandwidth) transducer array formed into a circular aperture with a 50-mm diameter. Each element was shaped into an arc in elevation to produce an acoustic focal length of 19 mm. The combined

![Fig. 1 Experimental setup of the oral-cavity illuminated photoacoustic computed tomography (OI-PACT) system: (a) Schematic of the OI-PACT system. (b) Detailed view of the optical fiber tip in the rat’s mouth. The origin of the z-axis was aligned with the plane 3 mm above the fiber tip.](https://nanophotonics.spiedigitallibrary.org/journals/Journal-of-Biomedical-Optics)
foci of all elements generated a central imaging region of 20-mm diameter and 1-mm thickness. Within the central imaging region, the system provided relatively uniform 0.10-mm radial (axial) resolution and <0.25-mm tangential (transverse) resolution.\textsuperscript{17} The center cross section of the ultrasonic transducer array determined the imaging plane, which was 3 mm above the fiber tip [Fig. 1(b)] when we imaged the brain base. The origin of the \( z \)-axis was aligned with the plane of the brain base. For the particular rat shown here, the elevational distance from the scalp to the brain base was around 13 mm.

The data acquisition was triggered by the laser’s Q-switch signal. After every other laser shot, the 8:1 multiplexers on the receiver boards forwarded the data from the transducer elements to the 64-channel acquisition board. A detailed description of the hardware can be found in previously published papers regarding the same data acquisition system.\textsuperscript{19,20} The impulse responses of the transducer elements were accounted for by deconvolving the raw channel data with the PA signal generated on the transducer surface. In order to reduce artifacts caused by the acoustic heterogeneities in the rat’s head, we used the simplified half-time image reconstruction algorithm,\textsuperscript{17,22} which backprojected only the first half of the received data.\textsuperscript{23}

To demonstrate the deep imaging capability provided by internal illumination, we imaged the deep brain structures of a healthy outbred rat (Hsd:Sprague Dawley® SD®, \( \sim \) 80 g). Since the Ti-Sapphire laser provided the strongest output at 780-nm wavelength, which happens to fall within the optical window, light at this wavelength was found to yield the highest quality images at depths. To investigate the advantages of internal illumination in deep brain imaging, after each measurement with fiber illumination, we also acquired corresponding control images with external illumination. Figure 2 shows a series of \textit{in vivo} images acquired over an elevational distance of 5 mm in the brain’s bottom region, with images acquired by internal illumination above [Figs. 2(a)–2(e)] and corresponding images obtained by external illumination below [Figs. 2(d)–2(f)]. We first coupled the light into the fiber for internal illumination to acquire an image [Fig. 2(a)] 1 mm below the brain base, i.e., \( z = -1 \) mm. We then flipped the mirror in front of the laser to redirect the light toward the top of the rat’s scalp to image the same brain layer with external illumination. The acquired control image is shown in Fig. 2(d). To image the shallower brain layers, we lowered the rat by 2 mm and acquired images with both internal and external illuminations, as shown in Figs. 2(b) and 2(e), respectively. Then, to image intermediate layers of the brain, we lowered the scanning stage in three increments of 1 mm each. In Fig. 2(c), the last one of such images acquired by fiber illumination, the recognizable structures include the thalamus, cerebral medulla, colliculus, cerebellum, and olfactory lobes. The corresponding control image is shown in Fig. 2(f). Clearly, for deep brain imaging, using the same PACT data collection system and image reconstruction algorithm, internal illumination provides much clearer images than external illumination, which barely showed any recognizable structures.

Figure 3(a) shows an \textit{in vivo} PA image of the rat brain base \( (z = 0 \) mm, ventral aspect). In the image, the left and right cerebral hemispheres, brain stem, hypothalamus, and anterior cerebral artery underneath the optic chiasma are clearly visible. In addition to the brain structures, the interface between the zygoma and the muscle tissue can also be seen. After the imaging experiment, we euthanized the rat in a CO\(_2\) chamber and then dissected its brain to photograph the anatomy in ventral view [Fig. 3(b)]. The PA image and photograph agree with each other despite the fact that the release from skull confinement slightly changed the shape of the cerebral hemispheres.

With pure endogenous hemoglobin contrast and 10 times averaging, each PA image shown above was acquired over 16 s. In principle, the system can acquire an image at each laser shot by using a matched 512-channel data acquisition system. In comparison with existing illumination schemes, internal illumination delivers more light into the bottom brain and enables deep brain imaging of live rats. With external illumination, current PA imaging systems limit themselves to the imaging of brain cortex and have not been able to image the deep brain structures of any animate animals.\textsuperscript{11–18}

In summary, by utilizing a fiber-transmitted internal illumination and full-view-array PACT system, we successfully
acquired cross-sectional images of the rat’s deep brain. The blood vessels under the brain, such as the anterior cerebral artery, as a part of the Circle of Willis, can potentially be investigated to study blood supply mechanisms and strokes caused by artery occlusion. Furthermore, the capability of bottom brain imaging also enables deep brain tumor studies. By implementing external and internal illuminations simultaneously, one can potentially achieve whole brain PA imaging. This illumination method can also be expanded to kidney and prostate imaging by inserting the fiber into the colon or to lymphonodus and heart imaging via fiber illumination from the esophagus. For human subjects, the sinuses can be potentially imaged by PACT with illumination from the oral cavity. Taking advantage of the wide choice of optical contrasts, such as near-infrared dyes and fluorescent proteins, different kinds of contrast mechanisms can be induced and potential benefit internally illuminated PACT for functional and molecular imaging.

Acknowledgments

The authors would like to thank Prof. James Ballard and Dr. Junjie Yao for close reading of the manuscript. This work was supported in part by National Institutes of Health Grants DP1 EB016986 (NIH Director’s Pioneer Award), R01 CA186567 (NIH Director’s Translative Research Award), R01 EB016963, and R01 EB010049. L.W. has a financial interest in Microphotoacoustics, Inc. and Endra, Inc., which, however, did not support this work.

References


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Journal of Biomedical Optics 016019-4 January 2015 • Vol. 20(1)

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